

DITERPENES OF *PODOCARPUS FERRUGINEUS* BARK

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Abstract—The following oxygenated diterpenes have been isolated from the bark of *Podocarpus ferrugineus* D. Don: isopimarol, isopimaric acid, sandaracopimaric acid, ferruginol, sugiol, sugiyl methyl ether, xanthoperol, royleanone, 6-dehydroroyleanone, cryptojaponol, 5 β -hydroxy-6-oxasugiyl methyl ether, 2-ketoferruginol and 2 β -acetoxysugiyl methyl ether.

INTRODUCTION

VARIOUS studies on the terpenic constituents of the New Zealand miro tree (*P. ferrugineus*) have led to the isolation of dipentene, limonene, α -pinene, cineole, cadinene, juniperol, rimuene, hibaene, phyllocladene, kaurene, ferruginol, 6-dehydroferruginol, sugiol, 5-dehydrosugiol and isopimaric acid.¹ In search of some supply of ferruginol we made an exhaustive survey of the terpene content of the bark. The hydrocarbons have been reported² to consist of longifolene, calamenene, α -copaene, α -muurolene, γ -muurolene, α -curcumene, α -bergamotene, β -selinene, the norditerpene 1a,³ isopimaradiene, and dehydroabietane, while an investigation of the oxygenated diterpenes and a discussion of their possible biosynthetic origin form the basis of the present communication.

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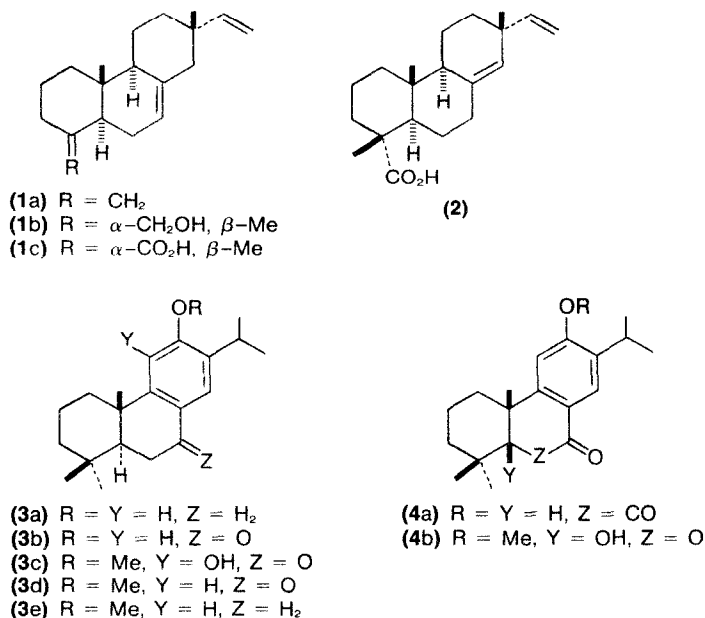
¹ HOSKING J. R. and SHORT, W. F. (1928) *Rec. Trav. Chim. Pays-Bas*, **47**, 834; HOSKING J. R., (1930) *ibid.*, **49**, 1036; BRANDT, C. W. and NEUBAUER, L. G. (1939) *J. Chem. Soc.*, 1031; (1940) 683; BRIGGS, L. H., CAWLEY, R. W., LOE, J. A. and TAYLOR, W. I. (1950) *ibid.*, 955; BROSSI A. and JEGE, O. (1950) *Helv. Chim. Acta*, **33**, 722; BRANDT C. W. and THOMAS, B. R. (1952) *Nature, Lond.*, **170**, 1018; BREDENBERG, J. B.-SON (1957) *Acta Chem. Scand.*, **11**, 932; BRIGGS, L. H., CAIN, B. F., CAMBIE, R. C. and DAVIS, B. R. (1960) *Tetrahedron Letters*, 18; BRIGGS, L. H. and CAMBIE R. C. (1960) *Tetrahedron*, **8**, 356; APLIN, R. T., CAMBIE, R. C. and RUTLEDGE, P. S. (1963) *Phytochemistry*, **2**, 205; BRIGGS, L. H., CAMBIE, R. C., RUTLEDGE, P. S. and STANTON, D. W. (1964) *Tetrahedron Letters*, 2223; DEL CASTILLO, J. B., BROOKS, C. J. W., CAMBIE, R. C., EGLINTON, G., HAMILTON, R. J. and PELLITT, P. (1967) *Phytochemistry*, **6**, 391; CAMBIE, R. C., MADDEN, R. J. and PARNELL, J. C. (1971) *Australian J. Chem.*, **24**, 217.

² KITADANI, M., YOSHIKOSHI, A., KITAHARA, Y., DE PAIVA CAMPELLO, J., MCCHESENEY, J. D., WATTS, D. J. and WENKERT, E. *Chem. Pharm. Bull. Tokyo*, **18**, 402.

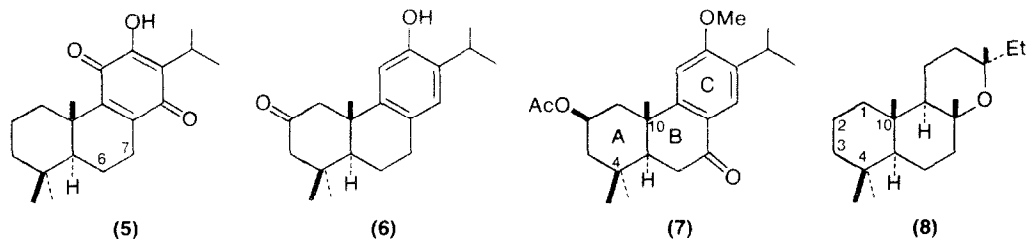
³ This hydrocarbon remained of unknown constitution.² Comparison of its infrared and pmr spectra with those reported recently for 1a [APPLETON, R. A. and ROERADE, J. (1969) *Chem. Commun.*, 1407], furnished kindly by Dr. Appleton, indicated the compounds' identity.

RESULTS AND DISCUSSION

The bark constituents isopimarol (1b), isopimaric acid (1c), sandaracopimaric acid (2), ferruginol (3a), sugiol (3b), sugiyl methyl ether (3d),⁴ xanthoperol (4a) and royleanone (5) were recognized by direct comparison with authentic samples. Two others, cryptojaponol (3c) and 5 β -hydroxy-6-oxasugiyl methyl ether (4b), were identified by partial synthesis⁵ and 6-dehydroroyleanone (6,7-dehydro-V) by conversion to royleanone on hydrogenation and subsequent air oxidation.⁶



The structures of two extractives, C₂₀H₃₀O₂ and C₂₀H₂₈O₃ substances, remained unelucidated, while those of two others, C₂₀H₂₈O₂ and C₂₃H₃₂O₄ substances, were found to be 6 and 7, respectively. The IR and PMR spectra of the C₂₀H₂₈O₂ compound limited it to a ketoferruginol system. While different from sugiol (3b) and hinokione, it yielded ferruginol on Wolff Kishner reduction. Thus it had to be 1-, 2- or 6-ketoferruginol. Its acceptance of four mass units on base-induced deuteration and reprotonation of its phenolic hydroxyl group revealed it as 2-ketoferruginol (6).⁷



⁴ SENGUPTA, P., CHOUDHURI, S. N. and KHASTGIR, H. N., (1960) *Tetrahedron*, **10**, 45.

⁵ WINKERT, E., MCCHESENY, J. D. and WATTS, D. J. (1970) *J. Org. Chem.*, **35**, 2422.

⁶ EDWARDS, O. E., FENIAK, G. and LOS, M. (1962) *Can. J. Chem.*, **40**, 1540.

⁷ The acetate of 6 has been reported as an intermediate in a study of the structure of salviol (2 α -hydroxyferruginol) [T. HAYASHI, T. HAYASHI, M. OHASHI, H. KAKISAWA, H.-Y. HSU and Y. P. CHEN, (1971) *Chem. Commun.*, 541].

The IR, UV and PMR spectra of the $C_{23}H_{32}O_4$ compound identified it as a ring A acetoxy derivative of sugiyl methyl ether (3d). Presence of the aromatic ketone moiety was confirmed by its hydrogenolysis and the consequent unmasking of a ferruginoid ring C PMR signal pattern. The multiplicity of the acetoxymethine signal indicated coupling to more than two neighboring hydrogens and the line shape was identical with that of the C(2)-H signal of the 2 β -acetoxy derivative of dihydromanoyl oxide (8). These facts showed the $C_{23}H_{32}O_4$ constituent to be 2 β -acetoxysugiyl methyl ether (7). The structure was corroborated by analysis of the $\Delta\delta$ values acquired by subtraction of the chemical shifts of the C(4) and C(10) methyl groups in 7, 7-deoxo-7 and all ring A acetoxyated dihydromanoyl oxides⁸ from those in 3d, ferruginyl methyl ether (3e) and 8, respectively (Table 1). The $\Delta\delta$ value for the natural compound and its 7-deoxo derivative agreed well with the value for 2 β -acetoxydihydromanoyl oxide.

TABLE 1. DIFFERENCES OF CHEMICAL SHIFTS OF METHYL GROUPS IN RING A ACETOXYLATED DITERPENIC COMPOUNDS FROM THOSE IN THEIR RING A UNOXYGENATED RELATIVES

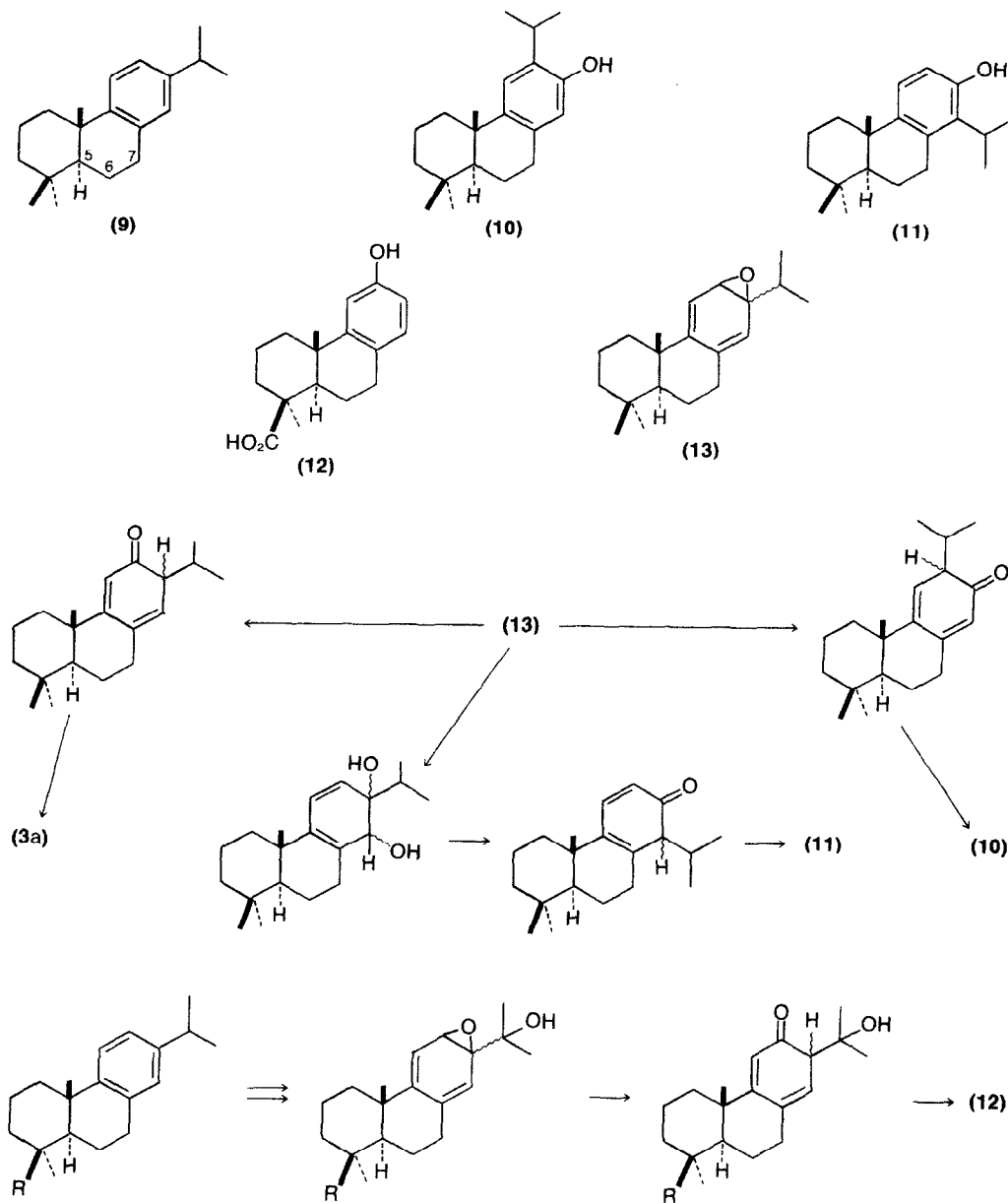
Compounds	$\Delta\delta$ (4 α -Me)	$\Delta\delta$ (4 β -Me)	$\Delta\delta$ (10-Me)
(1 α -Acetoxy-8) - (8)	0.05	0.04	0.06
(1 β -Acetoxy-8) - (8)	0.01	0.03	0.19
(2 α -Acetoxy-8) - (8)	0.06	0.09	0.10
(2 β -Acetoxy-8) - (8)	0.08	0.18	0.24
(3 α -Acetoxy-8) - (8)	0.02	0.08	0.03
(3 β -Acetoxy-8) - (8)	-0.01	0.06	0.04
(7) - (3d)	0.07	0.22	0.19
(7-Deoxo-7) - (3e)	0.07	0.18	0.19

The large array of oxygenated, ferruginoid substances in *P. ferrugineus* indicates a high activity of oxidizing enzymes in the plant. Perhaps the most significant finding in the survey of terpenic constituents of the miro tree is the discovery of dehydroabietane (9) as a natural product for the first time.² The appearance of this hydrocarbon, upon whose molecular skeleton the structures of all ring C aromatic diterpenic constituents of *P. ferrugineus* are based, lends a clue to the biogenetic origin of the naturally abundant, diterpenic phenols ferruginol (3a), semperviol (10) and totarol (11). In contrast to the earlier view of ferruginol being the first aromatic intermediate, from which totarol and podocarpic acid (12) are derived by acid-catalyzed isomerization and dealkylation, respectively,⁹ it is reasonable to assume now that the plant oxidases act upon dehydroabietane instead. Not only can oxidation be expected at the benzylic carbon 7 and subsequently at C(6) and C(5) (e.g. 3b-d and 4), but also oxidation on the benzene ring can be envisaged. Thus the natural phenols can be expected to be the end products of isomerization and hydration-dehydration processes on dehydroabietane oxide (13) or its oxygen position isomers.¹⁰ The biogenetic pathways are depicted in Scheme 1.

⁸ The authors are indebted to Professor P. Grant (University of Otago, Dunedin, New Zealand) for a gift of samples of these compounds.

⁹ WENKERT, E. and JACKSON, B. G. (1958) *J. Am. Chem. Soc.*, **80**, 211.

¹⁰ KASPEREK, G. J., BRUCE, P. Y., BRUCE, T. C., YAGI, H. and JERINA, D. M. (1973) *J. Am. Chem. Soc.*, **95**, 6041; KOBAL, V. M., GIBSON, D. T., DAVIS, R. E. and GARZA, A. (1973) *J. Am. Chem. Soc.*, **95**, 4420; ZIFFER, H., JERINA, D. M., GIBSON, D. T. and KOBAL, V. M. (1973) *J. Am. Chem. Soc.*, **95**, 4048; BRUCE, P. Y., KASPEREK, G. J., BRUCE, T. C., TAGI, H. and JERINA, D. M. (1973) *J. Am. Chem. Soc.*, **95**, 1673.



SCHEME 1.

EXPERIMENTAL

M.p.s were determined on a Reichert micro hot stage and are uncorrected. PMR spectra of CDCl_3 soln (unless otherwise noted) with internal TMS were recorded on Varian Associates A-60 and HA-100 spectrometers. ORD measurements were recorded with the use of a 1-cm quartz cell on a Durrum-Jasco ORD/UV-5 spectrophotometer. MS were determined in the direct inlet system of an AEI MS-9 spectrometer. Column chromatography was performed on G. F. Smith, 50-200 mesh silica gel or neutral, activity I Woelm alumina and TLC on Merck silica gel.

The previously described "acid fraction" from the extraction of *P. ferrugineus* D. Don bark² was acidified to pH 6 with 0.1 N HCl and extracted with CH_2Cl_2 (3×500 ml). The extract was dried and evaporated yielding 65 g of dark red gum. Furthermore, the alumina chromatography of the "neutral fraction" of the original bark extract was continued beyond the elution with light petrol.- Et_2O (9:1) which had led to a "hydrocarbon fraction" previously.²

Continuation of the elution of the alumina chromatographic column with petroleum-ether (4:1) gave 3.7 g of a mixture. Elution with a 3:2 solvent mixture yielded 63.7 g of ferruginol (3a) as viscous oil [acetate (Ac_2O -pyridine), colorless needles; m.p., m.m.p. $81-82^\circ$; $[\alpha]_D^{25} + 58^\circ$ ($c = 0.1$, MeOH)], with light petrol.- Et_2O (1:1) 46.5 g of a mixture, and with a 1:4 combination 9.5 g of a mixture. Chromatography of the 4:1 eluate on 150 g of silica gel and elution with light petrol. Et_2O (9:1) gave 1.6 g of oily ferruginol (3a), 600 mg of sugiyl methyl ether (3d); m.p., m.m.p. $136-137^\circ$; $[\alpha]_D^{25} + 32^\circ$ ($c = 0.1$, MeOH), and 20 mg of 2-acetoxysugiyl methyl ether (7); m.p. $168-170^\circ$; U.V. $\lambda_{\text{max}}^{\text{EtOH}}$ 230 nm (ϵ 17,500), 282 (14,500); IR (KBr) CO 5.76 (s), 6.00 (s) μ ; PMR δ 1.00, 1.21, 1.43 (s, 3, Me each), 1.21, 1.24 (d, 3, J 7 Hz, Me each), 2.10 (s, 3, Ac Me), 3.90 (s, 3, OMe), 4.69 (m, 1, OCH), 6.69 [s, 1, H(11)], 7.91 [s, 1, H(14)]; o.r.d. ($c = 0.05$, MeOH) $[\alpha]_D^{25} + 10^\circ$, $[\alpha]_{600} + 15^\circ$, $[\alpha]_{589} + 15^\circ$, $[\alpha]_{500} + 45^\circ$, $[\alpha]_{400} + 100^\circ$, $[\alpha]_{350} + 680^\circ$, $[\alpha]_{330} + 1100^\circ$, $[\alpha]_{318} 0^\circ$, $[\alpha]_{308} - 1200^\circ$, $[\alpha]_{300} - 1000^\circ$, $[\alpha]_{250} - 1000^\circ$, $[\alpha]_{240} 0^\circ$; m/e 372-2301 (calc. for $\text{C}_{23}\text{H}_{32}\text{O}_4$: 372-2300). Hydrogenation over Pd/C in EtOH gave a deoxy product spectrally identical with ferruginyl methyl ether in rings B and C; IR 6.00 μ CO band missing; PMR aromatic H signals shifted to 6.67 and 6.84 ppm.) Chromatography of the 1:1 eluate on 2 kg of silica gel and elution with light petrol. Et_2O (4:1) yielded 10.1 g of isopimarol (1b); m.p., m.m.p. $86-87^\circ$; $[\alpha]_D^{25} - 22^\circ$ ($c = 0.09$, MeOH), 150 mg of cryptojaponol (3c); m.p., m.m.p. $204-206^\circ$; $[\alpha]_D^{25} + 51^\circ$ ($c = 0.003$, MeOH), and 110 mg of a substance of unknown constitution; m.p. $210-212^\circ$; m/e 316-2038 (calc. for $\text{C}_{20}\text{H}_{28}\text{O}_3$: 316-2038). Chromatography of the 1:4 eluate on 500 g of silica gel and elution with light petrol. Et_2O (1:1) afforded 1.5 g of sugiyl methyl ether (3d) and 75 mg of a substance of unknown constitution; m.p. $180-182^\circ$; m/e 302-2247 (calc. for $\text{C}_{20}\text{H}_{30}\text{O}_2$: 302-2246).

Chromatography of the gummy "acid fraction" on 2 kg of silica gel and elution with light petrol. yielded 630 mg of an orange solid whose crystallization from heptane and sublimation gave 139 mg of 6-dehydroroyleanone (6,7-dehydro-5); m.p. $167-168^\circ$; spectra identical with those previously recorded.⁶ (Hydrogenation over Pd/C in EtOH yielded an unstable tetrahydro product whose air oxidation⁶ gave royleanone). Elution with light petrol.- Et_2O (9:1) led to 1.1 g of isopimarol (1b) and 5.2 g of light yellow gum whose crystallization from heptane afforded isopimaric acid (7c). Elution with 4:1 petrol.-ether yielded 1.2 g of ferruginol (3a) and 2.3 g of a yellow gum whose crystallization from heptane produced sandaracopimaric acid (7c); m.p., m.m.p. $171-172^\circ$; $[\alpha]_D^{25} - 25^\circ$ ($c = 0.13$, MeOH). Elution with a 2:1 solvent mixture gave 1.1 g of sugiyl; m.p., m.m.p. $283-284^\circ$ [acetate; m.p., m.m.p. $164-165^\circ$; $[\alpha]_D^{25} + 35^\circ$ ($c = 0.003$, MeOH)]. Elution with light petrol.- Et_2O (3:2) gave 248 mg of a pale yellow, amorphous solid whose crystallization from heptane-acetone yielded 25 mg of xanthoperol (4a); m.p., m.m.p. $265-270^\circ$ (sealed tube); $[\alpha]_D^{25} + 110^\circ$ ($c = 0.07$, MeOH). Elution with a 1:1 solvent pair produced 440 mg of a red oil whose rechromatography on 30 g of silica gel and elution with benzene led to 90 mg of golden plates of royleanone (5); m.p., m.m.p. $182-183^\circ$; $[\alpha]_D^{25} + 27^\circ$ ($c = 0.07$, MeOH). Elution with light petrol.- Et_2O (2:3) gave 470 mg of pale red oil whose titration with benzene and crystallization from heptane yielded 23 mg of 2-ketoferruginol (6); m.p. $232-234^\circ$; UV $\lambda_{\text{max}}^{\text{EtOH}}$ 227 nm (ϵ 8600), 284 (4300); IR (KBr) OH 2.91 (m), CO 5.85 (s), $\text{C}=\text{C}$ 6.20 (m) μ ; PMR δ 0.98, 1.11, 1.20 (s, 3, Me each), 1.23 (d, 6, J 7 Hz, Me_2), 6.58, 6.88 (s, 1, aromatic H each); o.r.d. ($c = 0.09$, MeOH) $[\alpha]_D^{25} + 17^\circ$, $[\alpha]_{600} + 33^\circ$, $[\alpha]_{589} + 50^\circ$, $[\alpha]_{500} + 117^\circ$, $[\alpha]_{400} + 233^\circ$, $[\alpha]_{350} + 500^\circ$, $[\alpha]_{320} + 1330^\circ$, $[\alpha]_{304} + 2470^\circ$, $[\alpha]_{290} 0^\circ$, $[\alpha]_{270} - 2220^\circ$, $[\alpha]_{260} - 2930^\circ$, $[\alpha]_{248} 0^\circ$, $[\alpha]_{238} - 3330^\circ$; m/e 300-212 (calc. for $\text{C}_{20}\text{H}_{28}\text{O}_2$: 300-209). Elution with a 1:4 solvent mixture gave 653 mg of a mixture of fatty acids, while elution with a 1:9 mixture led to 360 mg of a colorless, amorphous solid whose rechromatography on 25 g of silica gel, elution with benzene and crystallization from ethanol gave 46 mg of lactol 4b⁵ m.p. $168-169.5^\circ$; UV $\lambda_{\text{max}}^{\text{EtOH}}$ 222 nm (ϵ 5200), 270 (2600); IR (KBr) OH 3.00 (m), CO 5.89 (s), $\text{C}=\text{C}$ 6.24 (m) μ ; PMR δ 0.70, 1.19, 1.39 (s, 3, Me each), 1.22 (d, 6, J 7 Hz, Me_2), 3.92 (s, 3, OMe), 6.80 [s, 1, H(11)], 7.90 [s, 1, H(14)]; o.r.d. ($c = 0.42$, MeOH) $[\alpha]_D^{25} + 43^\circ$, $[\alpha]_{600} + 52^\circ$, $[\alpha]_{589} + 57^\circ$, $[\alpha]_{500} + 81^\circ$, $[\alpha]_{400} + 158^\circ$, $[\alpha]_{350} + 285^\circ$, $[\alpha]_{310} + 800^\circ$, $[\alpha]_{290} 0^\circ$, $[\alpha]_{270} - 80^\circ$, $[\alpha]_{248} 0^\circ$, $[\alpha]_{220} - 150^\circ$; m/e 332-202 (calc. for $\text{C}_{20}\text{H}_{28}\text{O}_4$: 332-199).

Deuteration and reduction of 2-ketoferruginol (6). A solution of 4 mg of 2-ketoferruginol (6) and 50 mg of NaOMe in 3 ml of D_2O was refluxed for 15 min and evaporated to dryness under vacuum. The procedure was repeated twice and the residue extracted with CH_2Cl_2 (3×25 ml). The extract was washed with 10 ml water and dried. Evaporation of the solvent gave 2 mg of material whose mass spectrum revealed strong suppression of the m/e 300 peak and appearance of a peak at m/e 304.

A solution of 4 mg of 2-ketoferruginol semicarbazone (m.p. $153-155^\circ$ from EtOH) and 200 mg of KOH in 5 ml of ethylene glycol was refluxed for 18 hr. It then was brought to pH 6 with 0.1 N HCl and extracted with 75 ml of CH_2Cl_2 $3 \times$. The extract was washed with 10% NaHCO_3 , dried and evaporated. A solution of the residual oil and 1 ml of Ac_2O in 1 ml of pyridine was kept at room temp. for 12 hr and then evaporated. Fractional sublimation of the residue yielded 1 mg of ferruginyl acetate, m.p. $80-82^\circ$, m.m.p. $79-82^\circ$.

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